



# A Commentary on “Outbreak of Cystoscopy-Related Urinary Tract Infections With *Pseudomonas aeruginosa* in South Korea, 2022: A Case Series”

Byoungkyu Han

Director of Urological Endoscopy Certification Committee, The Korean Urological Association Perfect Urology Clinic, Seoul, Korea

To the editor,

*Pseudomonas aeruginosa*, a Gram-negative bacterium commonly found in healthcare environments, is a major pathogen responsible for healthcare-associated infections (HAIs) and is recognized as a significant global public health concern. *P. aeruginosa* accounts for approximately 7%–10% of HAIs, causing a variety of infections, particularly ventilator-associated pneumonia, urinary tract infections (UTIs), and bloodstream infections [1-3].

Infections caused by *P. aeruginosa* lengthen hospitalization, increase medical costs, and, in some cases, are associated with high mortality rates [4]. A major concern is the bacterium's strong association with antibiotic resistance. Recent studies have reported multidrug resistance rates of up to 80.5% in clinical isolates, complicating treatment and posing significant challenges to infection control [5].

Hospital environments, especially moist areas such as sinks and faucets, serve as key reservoirs for *P. aerugi-*

*nosa* [6]. However, recent studies suggest that direct transmission from these environments may be lower than previously assumed, with some findings indicating that only 15% of bloodstream infections caused by *P. aeruginosa* are directly linked to environmental sources [4]. Therefore, a multifaceted approach—including rigorous environmental management, proper hand hygiene, antimicrobial stewardship, and continuous surveillance systems—is essential for the effective prevention and management of HAIs caused by this pathogen.

This study investigating *P. aeruginosa* infections following cystoscopy makes an important contribution to the field. The investigation involved 353 patients who underwent cystoscopy, of whom 6 developed febrile UTIs requiring hospitalization. *P. aeruginosa* was detected in 4 of these 6 patients, and environmental surveillance cultures identified the bacterium in a diluted solution sample of a dual-enzymatic detergent (EmPower, Metrex Research Corp., Orange, CA, USA). The authors reported that after changing the disinfectant and en-

**Corresponding author:** Byoungkyu Han

Director of Urological Endoscopy Certification Committee, The Korean Urological Association, Perfect Urology Clinic, 626 Gangnam-daero, Gangnam-gu, Seoul 06035, Korea

**Email:** [bkuro1@gmail.com](mailto:bkuro1@gmail.com)

<https://orcid.org/0000-0002-4862-5400>

**Received** February 3, 2025, **Revised** March 10, 2025, **Accepted** March 13, 2025



hancing the cleaning protocol, no further bacterial growth was observed, suggesting that the contaminated detergent was the likely source of the cystoscopy room associated infections [7].

Infection control in cystoscopy suites is critically important, particularly given the growing number of cystoscopic procedures. Beyond the limitations acknowledged by the authors, several key aspects require further discussion. Although *P. aeruginosa* was identified in the detergent, the precise route of contamination remains unclear. As this bacterium is a common opportunistic pathogen in healthcare settings, cross-transmission via healthcare workers' hands cannot be ruled out. Transmission of *P. aeruginosa* is known to occur more frequently in intensive care units (ICUs), and multiple outbreaks in neonatal ICUs have been reported. One study found that exposure to specific healthcare personnel was a significant risk factor for *P. aeruginosa* infection [8,9].

*P. aeruginosa* is a major cause of HAIs, particularly in moist environments where it forms biofilms that enable prolonged survival [10]. Preventing biofilm formation is crucial because biofilm-associated *P. aeruginosa* can persist under low-oxygen conditions and in extreme environments, making eradication difficult [11]. In healthcare settings, this bacterium is predominantly found on moist surfaces—including sinks, showerheads, and faucets—which serve as critical sources of HAIs [12].

Thus, stringent environmental management is essential for preventing *P. aeruginosa*-related HAIs. Key measures include:

(1) Hand hygiene and personal protective equipment (PPE) compliance – strengthen adherence to alcohol-based hand sanitizers, ensure the availability of proper handwashing facilities, and enforce mandatory PPE use in high-risk areas.

(2) Healthcare worker education – provide continuous training on disinfectant use, proper sterilization protocols, and infection control measures to prevent cross-contamination.

(3) Environmental cleaning and disinfection – imple-

ment routine cleaning and disinfection protocols, with particular attention to moist environments such as sinks, faucets, and showers, which serve as reservoirs for *P. aeruginosa*.

(4) Medical device reprocessing and sterilization – ensure strict adherence to proper cleaning, drying, and sterilization protocols for reusable medical devices, including cystoscopes, to prevent biofilm formation and bacterial persistence; rapid drying after cleaning is particularly important.

(5) Surveillance and monitoring systems – establish continuous environmental surveillance cultures and infection monitoring for early detection and intervention, particularly in high-risk areas such as hospital sinks, drains, and medical equipment storage units [10-13].

*P. aeruginosa* is generally susceptible to alcohol-based disinfectants, making alcohol-based hand sanitizers a crucial component of infection prevention. Notably, increased use of these sanitizers has been significantly correlated with a reduction in carbapenem-resistant *P. aeruginosa* infections [14]. However, some limitations exist; for instance, certain commercial alcohol wipes do not demonstrate sufficient bactericidal activity within 1–15 minutes of contact [15].

A critical survival mechanism of *P. aeruginosa* in hostile environments, including those with antibiotic exposure, is biofilm formation [10]. Therefore, preventing HAIs caused by this pathogen requires a dual approach—strict environmental management coupled with proper disinfection. Through these strategies, healthcare institutions can effectively inhibit biofilm formation and limit bacterial persistence and transmission. I would like to express our gratitude to the authors for conducting this pioneering study on cystoscopy-related *P. aeruginosa* infections in Korea. This research underscores the importance of infection control in endoscopic procedures and provides valuable insights for preventing similar outbreaks in the future.

• **Conflict of Interest:** The author has nothing to disclose.

## REFERENCES

1. Folic MM, Djordjevic Z, Folic N, Radojevic MZ, Jankovic SM. Epidemiology and risk factors for healthcare-associated infections caused by *Pseudomonas aeruginosa*. *J Chemother* 2021;33:294-301.
2. Volling C, Mataseje L, Graña-Miraglia L, Hu X, Anceva-Sami S, Coleman BL, et al. Epidemiology of healthcare-associated *Pseudomonas aeruginosa* in intensive care units: are sink drains to blame? *J Hosp Infect* 2024;148:77-86.
3. Álvarez-Lerma F, Olaechea-Astigarraga P, Palomar-Martínez M, Catalan M, Nuvials X, Gimeno R, et al. Invasive device-associated infections caused by *Pseudomonas aeruginosa* in critically ill patients: evolution over 10 years. *J Hosp Infect* 2018;100:e204-8.
4. Virieux-Petit M, Ferreira J, Masnou A, Bormes C, Paquis MP, Toubiana M, et al. Assessing the role of environment in *Pseudomonas aeruginosa* healthcare-associated bloodstream infections: a one-year prospective survey. *J Hosp Infect* 2024;156:26-33.
5. Asmare Z, Reta MA, Gashaw Y, Getachew E, Sisay A, Gashaw M, et al. Antimicrobial resistance profile of *Pseudomonas aeruginosa* clinical isolates from healthcare-associated infections in Ethiopia: a systematic review and meta-analysis. *PLoS One* 2024;19:e0308946.
6. Walker JT, Jhutti A, Parks S, Willis C, Copley V, Turton JF, et al. Investigation of healthcare-acquired infections associated with *Pseudomonas aeruginosa* biofilms in taps in neonatal units in Northern Ireland. *J Hosp Infect* 2014;86:16-23.
7. Kim B, Choi YS, Kang JK, Ha YS, Choi SH, Kim BS, et al. Outbreak of cystoscopy-related urinary tract infections with *pseudomonas aeruginosa* in South Korea, 2022: a case series. *Urogenit Tract Infect* 2024;19:97-103.
8. Jefferies JMC, Cooper T, Yam T, Clarke SC. *Pseudomonas aeruginosa* outbreaks in the neonatal intensive care unit--a systematic review of risk factors and environmental sources. *J Med Microbiol* 2012;61(Pt 8):1052-61.
9. Mayr A, Hinterberger G, Lorenz IH, Kreidl P, Mutschlechner W, Lass-Flörl C. Nosocomial outbreak of extensively drug-resistant *Pseudomonas aeruginosa* associated with aromatherapy. *Am J Infect Control* 2017;45:453-5.
10. Tuon FF, Dantas LR, Suss PH, Tasca Ribeiro VS. Pathogenesis of the *Pseudomonas aeruginosa* biofilm: a review. *Pathogens* 2022;11:300.
11. Qin S, Xiao W, Zhou C, Pu Q, Deng X, Lan L, et al. *Pseudomonas aeruginosa*: pathogenesis, virulence factors, antibiotic resistance, interaction with host, technology advances and emerging therapeutics. *Signal Transduct Target Ther* 2022;7:199.
12. Ambreetha S, Zincke D, Balachandar D, Mathee K. Genomic and metabolic versatility of *Pseudomonas aeruginosa* contributes to its inter-kingdom transmission and survival. *J Med Microbiol* 2024;73(2).
13. Browne K, Mitchell BG. Multimodal environmental cleaning strategies to prevent healthcare-associated infections. *Antimicrob Resist Infect Control* 2023;12:83.
14. Pires dos Santos R, Jacoby T, Pires Machado D, Lisboa T, Gastal SL, Nagel FM, et al. Hand hygiene, and not ertapenem use, contributed to reduction of carbapenem-resistant *Pseudomonas aeruginosa* rates. *Infect Control Hosp Epidemiol* 2011;32:584-90.
15. Tarka P, Chojecka A, Paduch O, Nitsch-Osuch A, Kanecki K, Kierzkowska A. Bactericidal activity of ready-to-use alcohol-based commercial wipes according to EN 16615 carrier standard. *Int J Environ Res Public Health* 2019;16:3475.